Molecular targeting of angiogenic blood vessels in atherosclerotic plaques with a new low molecular weight non-peptidic RGD mimetic



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INTRODUCTION

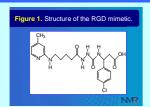
Atherosclerosis represents the leading cause of death and illness in developed countries and will soon become the pre-eminent health problem worldwide. Recent evidence suggests that neoanglogenesis greatly contributes to the instability of atherosclerotic plaque. The angiogenic blood vessels in the plaque may be particularly fragile and prone to micro-hemorrhage. The microvessels in the adventitia as well as in the plaque prominently express the integrin alpha(v)-beta(3), which is present at the sites of smooth muscle cell accumulation and angiogenesis in atherosclerotic plaques [1]. MRI is a promising technology that can provide noninvasive imaging with sub-millimeter resolution and high tissue contrast. In the present work, the noninvasive molecular imaging of plaque associated angiogenesis was assessed new low molecular weight non-peptidic RGD mimetic [2]. This alpha(v)-beta(3)-targeted molecule was grafted either to Gd-DTPA or to USPIO and the in vivo evaluation was performed on transgenic ApoE^{-/-} mice.

MATERIAL AND METHODS

Synthesis of mimRGD and of USPIO-g-mimRGD and Gd-DTPA-mimRGD

and Gd-DTPA-mimRGD

The RGD mimetic (Figure 1) was obtained as described by Sulyok et al [3]. The molecule was grafted onto DTPA by reaction with pSCN-Bz-DTPA (Macrocyclics, Dallas, Tx, USA); DTPA-mimRGD was then complexed with GdCl₃x6H₂O. For grafting onto magnetic nanoparticles (USPIO-g-mimRGD) the dextran coating of USPIO was previously treated with epichlorhydrin. Proton longitudinal (R₁) and transverse (R₂) relaxation rates were measured at 60 MHz and 37°C on a Bruker Minispec mq60 (Bruker, Karlsruhe Germany) Karlsruhe, Germany).



MATERIAL AND METHODS

In vivo MRI evaluation
To assess the molecula To assess the molecular imaging of atherosclerotic plaques, female C57Bl apoEm1Unc mice (8 and 18 months old, Charles Rivier Laboratories, Brussels, Belgium & St Germain sur l'Arbresle, France) were injected i.v. through the caudal vein with one of the following contrast agents: 0.1 mmol/kg b.w. of Gd-DTPA-mimRGD; 60 µmol/kg b.w of USPIO-g-mimRGD or of USPIO.

The MR images were acquired before and at different time intervals post-contrast up to 130 min, and at 24 h (for USPIO contrast agents). The animals were analyzed at 2 T (Oxford & SMIS imaging system) and at 4.7 T (Bruker AVANCE-200 system). For the images acquired at 2 T, a half-birdcage 200 system). For the images acquired at 2 T, a half-birdcage RF resonator was adapted to the size of the mouse (25-mm wide and 40-mm length) [3]. The microimaging device was used for the images acquired at 4.7 T. The following MRI protocols were used on the two imaging systems: (A) RARE, TR/TE = 1050/9.6 ms, RARE factor = 4, FOV = 23 mm, spatial resolution = 0.09 mm (4.7T); (B) RARE, TR/TE = 3000/20 ms, RARE factor = 4, FOV = 23 mm, matrix = 256, spatial resolution = 0.09 mm (4.7T); (C) fast SE, TR/TE = 2750/25-25 FOV = 23 mm, matrix = 256, spatial resolution = 0.09 mm (2T); (D) TOF 3D, TR/TE = 13/3 ms, flip angle = 40°, FOV = 40x20x40 mm, matrix = 256x128x64, spatial resolution = 0.156x0.156x0.625 mm (2T). Signal intensity (SI) values for each time delay were measured within several regions of each time delay were measured within several regions of interest in the arterial wall of the abdominal aorta by using the OSIRIS and IMDISP image analysis software. SI values were normalized with respect to a Gd-DTPA/saline solution in a test tube phantom. The data were averaged and the standard error was calculated for each time point. SI enhancement (\(\Delta S/N\)\%) was defined as the ratio of the difference between pre- and post-contrast SI to pre-contrast SI

Blood pharmacokinetics and validation of the

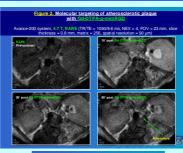
specific interaction with integrins

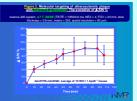
The specific targeting of integrins was tested on Jurkat cells stimulated with phorbol myristate acetate (PMA). Plasma pharmacokinetics of USPIO-g-mimRGD were assessed on Wistar rats, healthy or with hepatitis induced by concanavalin

RESULTS

The in vivo molecular imaging of atherosclerotic plaques: Gd-DTPA-mimRGD

The images obtained at 4.7 T with Gd-DTPA-mimRGD have shown an important enhancement of the signal at the level of atherosclerotic plaque in abdominal aorta 80 min post-contrast (Figure 2). The evolution of $\Delta S/N\%$ (Figure 3) measured on 10 ROI that cover 8 mm along the abdominal aorta of an ApoE[±] mouse indicates the constant enhancement of SI, which attained a value of 103% 80 min after the administration of Gd-DTPA-mimRGD.

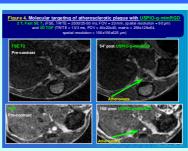


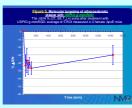


RESULTS

The in vivo molecular imaging of atherosclerotic plaques: USPIO-g-mimRGD

Figure 4 shows the images of the atherosclerotic plaque obtained at 2T, 54 min (FSE T_2) and 160 min (TOF) post USPIO-g-mimRGD. The decrease of SI indicate an accumulation of USPIO-g-mimRGD at the level of the atherosclerotic plaque, mainly around the blood vessel lumen. The estimation of $\Delta S/N\%$ on the FSE T₂ images (Figure 5) evidenced that SI decreased down to - 44%, 34 min post-contrast, the lowest level being observed 130 min post-contrast. The SI tended to return to the baseline





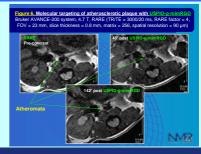
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RESULTS

The in vivo molecular imaging of atherosclerotic plaques: USPIO-g-mimRGD

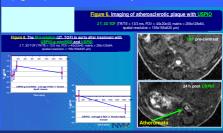
The images obtained at 4.7 T with RARE protocol (Figure 5) are comparable to those obtained at 2 T by using FSE T2 (Figure 2), meaning that SI changed around the blood vessel lumen giving the appearance of



RESULTS

The in vivo imaging of atherosclerotic plaques: USPIO

USPIO (2T) did not produce any change of SI at the level of atherosclerotic plaque during the subsequent 2 h post-contrast. A black strip was observed on TOF images at The lumenas. A Brack strip was observed on TOP images at the lumenal side of the blood vessel wall 24 h post-contrast (Figure 7). The estimation of ΔS/N% on TOF images (Figure 8) confirms this type of SI evolution, which is probably correlated to USPIO uptake by macrophages. By the contrary, USPIO-g-mimRGD produced the decrease of ΔS/N% on TOF images (Figure A) down to 40% 44 miles (Figure A) down to 40% 44 mi (Figure 4) down to - 39% 24 min post-contrast.



RESULTS

USPIO-g-mimRGD: validation of the specific interaction with integrins and pharmacokinetic characterization

specific interaction of USPIO-g-mimRGD with integrins expressed by Jurkat cells was evaluated by a new technique, C-MALISA [4]. The \mathbf{K}^*_{d} of this interaction was estimated to 1.13 x10-8 M. A diminished blood clearance in pathological conditions as compared to different controls (healthy animals treated with USPIO-g-mimRGD, animals treated with USPIO) was observed for USPIO-g-mimRGD: prolonged half-life of elimination (266 min); diminished total clearance (0.235 mL/kg/min); diminished VDss (0.066 L/kg).

CONCLUSIONS

The results suggest that integrin targeting with mimRGD-grafted contrast agents contributes to the high-resolution *in vivo* molecular imaging of unstable atherosclerotic lesions. The new compounds could help to the detection of various pathologies associated with angiogenic blood vessel development, such as the cancer

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